Identification of Weed hosts of Rice yellow mottle Sobemovirus at Sayen Gobirawa, Northern Nigeria

¹Salaudeen, M.T., ²Banwo, O.O., ²Kashina, B. D. and ²Alegbejo, M. D. Department of Crop Production, Federal University of Technology, P.M.B. 65, Minna, Nigeria; E-mail: mtsalaudeen@vahoo.co.uk; Tel.: (+234) 08063330183 ²Department of Crop Protection, Ahmadu Bello University, P.M.B. 1044, Zaria, Nigeria

Abstract: Indew nities redmun teel

Liller size A study was carried out to identify the weed hosts of Rice yellow mottle Sobemovirus (RYMV) at Sayen Gobirawa, northern Nigeria during the 2005/06 cropping season. Weed species with or without the typical symptoms of the virus were collected in and around the RYMV-infected rice field. The detection of the pathogen was based on double antibody sandwich enzyme - linked immunosorbent assay (DAS-ELISA) and bioassays of cultivated plants. All the weed species (Cynodon dactylon (L.) Pers, Cyperus esculentus L., Cyperus rotundus L., Eleocharis complanata Boeck, Eleusine indica (L.) Gaertner, Fuirena umbellata Rottb., Imperata cylindrica L., Kyllinga pumila Michaux and Paspalum vaginatum Sw.) tested positive for RYMV. The present results suggest that some weed grasses belonging to the families Cyperaceae and Poaceae could serve as natural hosts of RYMV and therefore could play a significant role in the spread of the disease. conference of the African Small Ruminant Research Network, Arusha, Tanzania, 7-11

Keywords: Weed hosts; Rice yellow mottle Sobemovirus; Sayen Gobirawa; Nigeria Sayen Gobirawa; Nigeria

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INTRODUCTION

Rice yellow mottle Sobemovirus is positive sense RNA virus (Bakker, 1970; 1974). The virus was first reported in 1966 at Otonglo, Kenya, near Lake Victoria in East Africa (Bakker, 1970). It was later found in West Africa (Raymundo and Buddenhagen, 1976), southern Africa and Madagascar (Reckhaus and Randrianangaly, 1990). In 2001, it was noticed in Central Africa (Traore et al., 2001) and by 2002 it was already in Europe (Koklu and Yilmaz, 2004). The virus is spreading rapidly between and within the countries of the world.

Infection of rice by RYMV is becoming increasingly important in Nigeria too (Abo et al., 2002). Following its first appearance in 1978 at Ibadan in Oyo State (IITA, 1979) and spread to other parts of the country (Awoderu, 1991; Singh et al., 1997; Abo et al., 2002; Alegbejo et al., 2006), the incidence ranges from 5 to 100 % (Rossel et al., 1982; Awoderu, 1991; Alegbejo et al., 2006). Consequently, yield losses averaging 25 to 100 % have been recorded (Rossel et al., 1982; Alegbejo et al., 2006). Various weed species have been implicated in the epidemiology of the virus. Bakker (1974) reported that the pathogen was probably on grass weeds prior to rice cultivation and then spread to it. Additionally, Okioma et al. (1983) and Awoderu (1991) reported that some wild grass species which occur abundantly around rice fields are important reservoirs of the pathogen. Earlier studies reported that RYMV has a narrow host range which is restricted to species in the Poaceae, tribes Oryzae and Eragrostidae (Bakker, 1974). Experimental hosts of the virus are the Cynodon dactylon (L.) Pers., Digitaria sanguinalis (L.) Scop., Dinebra retroflexa (Vahl) Ponzer, Echinochloa colona (L.) Link and Eleusine indica (L.) Gaertner (Bakker, 1974; Okioma et al., 1983; Awoderu, 1991; Konate et al., 1997. Abo et al., 2003). On the other hand, Ischaemum rugosum Salisb and Oryza longistaminata A. Chev and Roehr have been reported as its natural weed hosts (Bakker, 1974; Awoderu, 1991; Konate et al., 1997; Abo et al., 2002). Knowledge of weed hosts is essential for the characterization of virus, detection of new strains and sustainable management strategies. The major objective of this study was to identify the weeds that serve as natural reservoirs of RYMV.

Materials and Methods

Materials and methods
Field surveys and samplings

Weeds with symptoms such as leaf mottling yellowing, blistering, curling, narrowing distortion, vein clearing and stunting were collected randomly from Sayen Gobirawa (11°1'N, 7°40'E, 650 m above sea level). Weeds without symptom of infection were also collected. The samples were packed in plastic bags, labeled and stored in the freezer (-20 °C) at the Virology Laboratory of the Department of Crop Protection, Ahmadu Bello University, Zaria, until used. Samples were collected from April, 2005 to June, 2006. They were taken from the University, Zaria, until used. Camples 11.00 line weed species were studied. I new were taken from the rice field, edges and vicinity of the RYMV-infected plants. For each weed species 36 symptomatic and symptomless samples were collected. In all, nine weed species were studied.

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Serology

The serological detection of the virus was based on double antibody sandwich enzyme linked immunosorbent assay (DAS ELISA) as described by Clark and Adams (1977). Each weed species was evaluated in one homogenate, in duplicate wells of the polystyrene microtitre plate. The monoclonal antibody against RYMV (AS 0478/11) supplied by Dr. S. Winter of the Plant Virus Collection Centre, DSMZ -Braunschweig, Germany was used. The Oyo RYMV isolate (DSMZ 0478) was used as a positive control while leaf extract of healthy non cereal plant (*Tridax procumbens* L.) served as negative control. The various buffers used include coating buffer (1.5 g sodium carbonate, 2.93 g sodium bicarbonate, 0.20 g, sodium azide pH 9.6, conjugate buffer (PBS-T, 2 % PVP + 0.2 % egg albumin per litre) and substrate buffer (97 ml diethanolamine, 0.2 g sodium azide per H₂O, pH 9.8). The experiment was carried out twice.

Bioassay tests

Crude extract from symptomatic and symptomless samples was prepared separately by homogenizing leaves with 0.1M phosphate buffer, pH 7.4 in 1:10 ratio (w/v). Carborundum powder (600 mesh) was added to the extract of each sample and the homogenate rubbed onto leaves of 2 week old six seedlings of the rice cultivar Bouake 189, which is highly susceptible to RYMV. Three seedlings of each species were left uninoculated to serve as controls. The test plants were kept in a screen house at 22 30 °C and inspected for symptoms daily. A total number of 6 observations were made at weekly interval. symptomiless hosts at a virus car

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unknown since bloassay has to be carried out in each to confirm ratio presence. Therefore,

1970). Bakker (1974) stated that perennial weeds plovide avenues for the virus to sur-

Back inoculation

The leaves of plant species that remained symptomless were back assayed on Bouake 189 seedlings and observed for 4 weeks. Three uninoculated seedlings served as controls.

Results and Discussion

Colourless

Reaction of the weed samples in serology and bioassay tests 1

The entire weed species tested positive for RYMV while the healthy controls were negative (Table 1). When the plants were inoculated with extract of symptomatic samples none of the weed species induced completes susceptibility. A susceptibility of 50 % (3/6) resulted from the extract of the weed species Cyperus esculentus L., C. rotundus L., Fuirena umbellata Rottb., and Kyllinga pumila Michaux within the family Cyperaceae. In the family Poaceae, the weed species Cynodon dactylon (L.) Pers, Eleusine indica (L.) Gaertner and Imperata cylindrica L. induced 50 % infectivity. Conversely, Eleocharis complanata Boeck and Paspalum vaginatum Sw. resulted in a susceptibility of 33.3 % (2/6) within the Cyperaceae and Poaceae family, respectively (Table 2). When the plants were treated with extract of symptomless samples I. cylindrica induced 100 % (6/6) infection while a susceptibility of 83.3 % (5/6) was induced by E. indica (Table 2). Infectivity of 66.7 % (4/6) resulted from C. dactylon, F. umbellata, K. pumila and P. vaginatum while the weed species C. esculentus, C. rotundus, and E. complanata produced 50 % infection (Table 2). When the result of the bioassay test was considered in general terms, 51.7 % (31/60) members of the Cyperaceae showed the typical symptoms of RYMV. In contrast, 62.5 % (30/48) species of the Poaceae induced the disease. In all the back inoculation tests the indicator plants exhibited the typical symptoms of the virus.

Table 1: Reaction of the various weed species in double antibody sandwich enzyme linked immunosorbent assay Sero-reaction

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Reaction of the weed samples in serology and bioassay tests 2

All the weed species tested positive for the virus (Table I). In the bioassay tests it was observed among the members of Cyperaceae that the level of infection 66.7 % (4/6) produced by *K. pumila* was the highest when the plants were inoculated with extract of symptomatic samples (Table 2). Fifty % susceptibility resulted from inoculation with extract of *C. esculentus*, *E. complanata*, and *F. umbellata*. On the other hand, susceptibility of 33.3 % was produced by *C. rotundus*. Among the species of Poaceae inoculation of the test plants with symptomatic samples resulted in complete infection by extract of *I. cylindrica*. On the other hand, susceptibility of 66.7, 50.0 and 33.3 % each was recorded upon inoculation with *C. dactylon*, *E. indica*, and *P. vaginatum*, respectively. When the plants were treated with extract of symptomless samples the members of Cyperaceae C. esculentus, *F. umbellata*, and *K. pumila* induced the highest (83.3 %) susceptibility while *C. rotundus* and *E. complanata* produced 66.7 % infection (Table 2). Within the members of Poaceae, inoculation with extract of *C. dactylon* resulted in the highest (83.3 %) level of susceptibility. However, the use of *E. indica*, *I. cylindrica* and *P. vaginatum* produced 66.7 % infection each (Table 2). Considering the level of infection in general terms 63.3 % (38/60) members of the Cyperaceae induced RYMV symptoms while 66.7 % (32/48) species of the Poaceae produced the disease in the test plants (Table 2).

The detection of the virus in all the weed species indicates their importance in the ecology and survival of the pathogen. Additionally, the ability of the virus extract from the symptomless weed samples to induce RYMV symptoms suggests their significant role in the epidemiology of the disease. Rosenkranz (1983) reported that symptomless hosts of a virus can be particularly troublesome if they act as overwintering hosts and remain unknown since bioassay has to be carried out in order to confirm their presence. Therefore, timely and effective control of the weeds is essential. The susceptibility of all the perennial weed species is of special interest since they are capable of harbouring the pathogen all the year round and for extended number of years. The role of the susceptible annual weeds is also of great concern since the virus can survive in infected dry leaves (Bakker, 1970). Bakker (1974) stated that perennial weeds provide avenues for the virus to survive during the off season and then serve as sources of inocula at the beginning of the new season. Additionally, the annual weeds could also harbour the virus during the growing season and serve as sources of inocula for secondary spread (Rosenkranz, 1980). These were probably responsible for the high RYMV incidence (100 %) observed during

the field surveys (data not shown).

The detection of RYMV in mechanically inoculated *Eleusine indica* (L.) Gaertner bas been reported by Awoderu (1991) while this study shows that natural infection is also possible. However, this report is not in agreement with the earlier work of Abo *et al.* (2003), who reported it as a non-host of RYMV. This might be due to the difference in the pathogenicity of the isolates involved (Fargette *et al.*, 2000; N'Guessan *et al.*, 2001). Also, the detection of the virus in *Imperata cylindrica* contrast to the findings of Abo *et al.* (2003), could also be attributed to the difference in the pathogenicity of the isolates (Fargette *et al.*, 2001; N'Guessan *et al.*, 2001) evaluated. The report of RYMV in the mechanically inoculated *Cynodon dactylon* was earlier reported by Awoderu (1991) while this study reports natural infection by the pathogen. The differences observed in the symptoms expressed upon inoculation with extract of the various weed species could be attributed to the difference in the virulence (N'Guessan *et al.*, 2001) of the strains of the virus invading the weeds.

Table 2: Reaction of Bouake 189 rice plants to extract of weed samples collected from RYMV infected field at Sayen Gobirawa in 2005/2006

Weed species	EXPERIMENT I										EXPERIMENT 2									
		Number of plants infected when inoculated with extract of:				Back inoculation				Nu	Number of plants infected when inoculated with extract of:					Back inoculation				
	LC	ST	C	SL	C	ST	С	SL	С	ST	С	SL	С		ST	C	CI	e C		
Cyperaceae						0.80			Mar. I				<u> </u>	-	31	C	SL	C		
Cyperus esculentus	P	3/6	0/3	3/6	0/3	4/6	0/3	5/6	0/3	3/6	0/3	5/6	0/3		odpo	0.10	System System	COL		
C. rotundus	P	2/6	0/3	3/6	0/3	3/6	0/3	4/6	0/3	2/6	0/3	4/6	0/3	must	5/6	0/3	6/6	0/3		
Eleocharis complanata	P	2/6	0/3	3/6	0/3	3/6	0/3	3/6	0/3	3/6	0/3	4/6			3/6	0/3	5/6	0/3		
Fuirena umbellata	P	3/6	0/3	4/6	0/3	3/6	0/3	4/6	0/3	3/6	0/3		0/3		3/6	0/3	3/6	0/3		
Kyllinga pumila	P	3/6	0/3	4/6	0/3	4/6	0/3	4/6	0/3	4/6	0/3	5/6	0/3		3/6	0/3	6/6	0/3		
Poaceae										470	0/3	5/6	0/3		5/6	0/3	5/6	0/3		
Cynodon dactylon	P	3/6	0/3	4/6	0/3	3/6	0/3	5/6	0/3	4/6	0/3							0/3		
Eleusine indies	A	3/6	0/3	5/6	0/3	4/6	0/3	5/6	0/3	3/6	0.00	5/6	0/3		4/6	0/3	6/6	0/3		
Imperata cylindrical	P	3/6	0/3	6/6	0/3	3/6	0/3		.,,	6/6	0/3	4/6	0/3	10	3/6	0/3	5/6	0/3		
Paspalum vaginatum	P	2/6	0/3	4/6	0/3	3/6	0/3	3/6	0/3	2/6	0/3	4/6	0/3	111133	barron.	BOY F	4/6	0/3		
A = Annual species;	P = P	erennia	al spe	cies; C	= Con	trol; L	C = Li	fe Cv		= Sympt	0/3	4/6	0/3		3/6	0/3	5/6	0/3		

Therefore, these weeds could serve as sources of inocula in immunological and molecular characterization studies aimed at identifying the strains of the virus (Rosenkranz, 1987) as well as their distribution. Interestingly, inocula could also be obtained from these weed species by plant breeders to confer resistance on the RYMV susceptible rice cultivars.

Although it has been reported that weed hosts of RYMV are mainly in the Poaceae family, (Bakker, 1974) RYMV is highly variable so that biological and serological differences among its isolates are not uncommon (Konate et al., 1997; N'Guessan et al., 2000, 2001; Pinel et al., 2000). This was probably responsible for infection of the weed species belonging to the family Cyperaceae. Lake Victoria başin in Kenya, Trop. Peşt Manage, 29: 295-296

Conclusion

The results of this study indicated that weed species in the family Cyperaceae could also harbour the virus. However, this is the first report of RYMV on Cyperus esculentus, Cyperus rotundus, Eleocharis complanata, Fuirena umbellata, Imperata cylindrica, Kyllinga pumila, and Paspalum vaginatum. Future natural weed hosts studies should evaluate plant species in other families. Will happen happen

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- References departe asional box assign Abo, M.E., Ukwungwu, M.N. and Onasanya, A. (2002). The distribution, incidence, natural reservoir hosts and insect vectors of Rice yellow mottle virus (RYMV), Genus Sobemovirus in northern Nigeria. Tropic., strains A and B and Sugarcana mosaic 20(4): 189 - 202.
- Abo, M.E., Alegbejo, M.D., Sy, A.A., Adeoti, A. A. and Marley, P.S. (2003). The host range of Rice yellow mottle virus, Genus Sobemovirus in Cote d'Ivoire. Samaru J. Agric. Res. 19: 69-78.
- Alegbejo, M.D., Raji, B.A., Abubakar, I. U. and Banwo, O.O. (2006). Rice yellow mottle virus disease, a new disease of rice in Zamfara, Nigeria. Int. Rice Res. Notes, 31:1. 668 608. Quillogs mobilish nee growing areas of Nigeria
- Awoderu, V. A. (1991). Rice yellow mottle virus in West Africa. Trop. Pest Manage., 37(4): 356 362.
- Bakker, W. (1970). Rice yellow mottle virus, a mechanically transmissible virus disease of rice in Kenya. Neth. J. Plant Pathol., 76:53-63.
- Bakker, W. (1974). Characterization and ecological aspects of Rice yellow mottle virus in Kenya. [Ph.D Thesis] Agricultural University, Wageningen, The Netherlands, 2008 (2016)
- Clark, M.F. and Adams, A.N. (1977). Characteristics of the microplate method of enzyme-linked immunosorbent assay for the detection of plant viruses. J. Gen. Virol., 34: 475-483.
- Fargette, D., Pinel, A., Albar, L. Sadiky, R., N'Guessan P., Corgier S. Frutois R. Nottenghem. J.L., and Ghesquiere A. (2001). Assessment of biological, serological and molecular Noπengnem. J.L., and Rice yellow mottle virus isolates from different geographical areas, p. 91. In: Sy variability of a range of Rice yellow mottle virus (PVM). Feet geographical areas, p. 91. In: Sy variability of a range of Allo A. Rice yellow mottle virus (RYMV): Economic importance, diagnosis and A.A., Hughes J. and Diallo A. Rice yellow mottle virus (RYMV): Economic importance, diagnosis and A.A., Hugnes J. and Diallo A.A., Hugnes J. and D
- IITA (International Institute of Tropical Agriculture). (1979). International Institute of Tropical Agriculture Annual Report for 1978, Ibadan, Nigeria .108:20 21.
- Koklu, G. and Yilmaz, O. (2004). Research on Rice ragged stunt and Rice yellow mottle viruses on rice grown in Edirne, Turkey. Cereal Res. Comm., 32(3): 387-395.
- Konate, G., Traore, O. and Coulibaly, M. (1997). Characterization of *Rice yellow mottle virus* isolates in Sudano-Sahelian areas. Arch. Virol., 142: 1117-1124.

- N'Guessan, P., Pinel, A., Caruana, M., Fruitos, R., Sy, A. Ghesquiere, A. and Fargette, D. (2000). Evidence of the presence of two serotypes of *Rice yellow mottle Sobemovirus* in Cote d'Ivoire. Eur. J. Plant Pathol, 106: 167-178.
- N'Guessan, P.N., Pinel, A., Sy, A. A., Ghesquiere, A. and Fargette, D. (2001). Distribution, pathogenicity, and interactions of two strains of *Rice yellow mottle virus* in forested and savanna zones of West Africa. Plant Dis., 85(1): 59-64.
- Okioma, S. N. W., Muchoki, R. N. and. Gathuru, E. M. (1983). Alternative hosts of *Rice yellow mottle virus* in the Lake Victoria basin in Kenya. Trop. Pest Manage., 29: 295-296.
- Pinel, A., N'Guessan, P. N., Bousalem, M., and Fargette, D. (2000). Molecular variability of geographical distinct isolates of *Rice yellow mottle virus* in Africa. Arch. Virol., 145: 1621-1638.
- Raymundo, S.A. and Buddenhagen, I.W. (1976). A virus disease in West Africa. Int. Rice Commiss. Newsl., 25:58.
- Reckhaus, P.M. and Randrianangaly, S. (1990). Rice yellow mottle virus (RYMV) on rice in Madagascar. Int. Rice Res. Notes, 15 (1): 30.
- Rosenkranz, E. (1980). Taxonomic distribution of native Mississippi grass and species susceptible to Maize dwarf and Sugarcane mosaic viruses. Phytopathol., 70: 1056-1061.
- Rosenkranz, E. (1983). Susceptibility of representative native Mississippi grasses in six subfamilies to Maize dwarf mosaic virus strains A and B and Sugarcane mosaic virus strain B. Phytopathol., 73:1314 1321.
- Rosenkranz, E. (1987). New hosts and taxonomic analysis of the Mississippi native species tested for reaction to Maize dwarf mosaic and Sugarcane mosaic viruses. Phytopathol., 77 (4): 598-606.
- Rossel, H. W., Thottappilly, G. and Buddenhagen, I.W. (1982). Occurrence of *Rice yellow mottle virus* in two important rice growing areas of Nigeria. FAO Plant Protect. Bull., 31: 137-139.
- Singh, B.N., Fagade, S., Ukwungwu, M.N., Williams, C., Jagtap, S.S., Oladimeji, O., Efisue, A. and Okhidievbie, O. (1997). Rice growing environments and biophysical constraints in different agro-ecological zones of Nigeria. Met. J., 2(1): 35-44.
- Traore, O., Pinel, A., Fargette, D. and Konate, S. (2001). First report and characterization of *Rice yellow mottle* virus in Central Africa. Plant Dis., 85:920.

Clark, M. F. and Adams, A. N. (1977). Characteristics of the micropiate method of engage in ked unusupsort and

ATA (International Institute of Tropical Agriculture), (1979), International Institute of Tropical Agriculture Annual Report for 1978, Ibaden, Nigeria, 198:20-21.

foldu, G. and Yilmaz, O. (2004). Research on Rice ragged stunt and Rice yellow mottle viruses on rice grown in

Konste, G., Tragre, O. and Coulibrity, M. (1997), Characterization of Rice yellow mortle virus isolates in Sugano-

Nottenghem. J.L., and Chesquiere A. (2001) Assessment of biological, serological and molecular variability of a range of Rice yellow mottle virus isolates from different geographical areas, p. 91 in: Sy A. Hughes J. and Diallo A. Rice yellow mottle virus (RYMV). Economic importance, diagnosis and management strategies. Bouake, Cote d'Ivoire, West Ainca Rice Development Association, p. 252.

Sadiky, R. N'Guessan P., Corgier S. Frutois R.

assay for the detection of plant viruses. J. Gen. Virol., 34: 475-483.

Edirne, Turkey, Cereal Res. Comm., 32(3): 387-395.

Sanctian areas, Arch. Virol., 142, 1117-1124