POTENTIALS OF YEAST ISOLATES FOR BIOSURFACTANT PRODUCTION

₁₀yeleke, S. B., ¹Goro, M. A., *¹Oyewole, O. A., ²Okeke, K. S., ³Ayisa, T. T., ⁴Mohammed, S. S. D.

appendent of Microbiology, Federal University of Technology, Minna, Nigeria apepartment of Nutrition and Dietetics, Federal Polytechnic Bida, Nigeria apepartment of Biological Sciences, Federal Polytechnic Bida, Nigeria ³pepartment of Microbiology, Kaduna State University, Kaduna State, Nigeria

ABSTRACT

Twenty yeast isolates were screened for potential to produce biosurfactants using three Twenty methods namely: collapse of oil drops, oil spreading or displacement and emulsification potential. Of the twenty yeast isolates screened, sixteen (80.0 %) showed collapse of oil drops. Eleven isolates (55 %) were able to displace the oil with Saccharomyces cerevisiae H02, Candida boleticola H09 and Rhodotorula bogoriensis H15 having the highest diameter of displacement of 6.2 cm, 5.3 cm and 4.7 cm respectively. The isolates had emulsifying capacities that ranged from 6.67 % to 33.33 %. The results showed that Saccharomyces cerevisiae H02, Candida boleticola H09 and Rhodotorula bogoriensis H15 were considered efficient potential candidates for biosurfactants production. Keywords: yeast, biosurfactants, oil collapse, oil displacement, emulsification

*Correspondence author: oa.oyewole@futminna.edu.ng

INTRODUCTION

amphiphilic · Biosurfactants secreted which are compounds, extracellularly by microorganisms (Luna et al., 2013). They can be used in many industries as bio-emulsifiers in the food industries; as antimicrobial agent; in detergent formulation; in bioremediation processes as well as in enhanced crude oil recovery. Even though yeasts are known for producing biosurfactants in higher concentrations than bacteria, which is an advantage regarding microorganisms, minimal studies have reported biosurfactants synthesized by yeasts (Hua et al., 2003). Hence the need biosurfactant to explore in yeasts production, which was the basis for this study.

MATERIALS AND METHODS

Yeast strains

Twenty yeast isolates were used. The yeasts were obtained from Department of University Federal Microbiology, Technology, Minna, Nigeria. Drop collapse test: Oil drop collapse assay

developed by Jain et al. (2011) was adopted. oil

The test: displacement displacement test was done according to methods described by Rodrigues et al.

Emulsification (2006)activity: Emulsification towards biosurfactants kerosene was studied using methods of Youssef et al. (2004).

Statistical analysis: Data was analyzed statistically using error mean square and correlation analysis. Computer statistical package SPSS 9.0 was used.

RESULTS

Of the twenty yeast isolates screened, sixteen (80.0 %) showed positive result (that is, dispersed the oil around the

liquid droplet). Of the sixteen positive isolates, only seven (43.75 %) were able to collapse the oil droplet (that is, caused it to spread out and appeared flat on the solid surface within one minute. The results of the oil drop collapse caused by yeast isolates are shown in Table 1

Table 1: Extent of collapse of oil drops caused by yeast isolates

Coded yeast isolates	Reaction	
Pochia strasburgensis H01*	+	
Saccharomyces cerevisiae H02	++	
Saccharomy ces cerevisiae H03	++	
Cryptococcus laurentii H04	+	
Cryptococcus skinneri H05	+	
Yarrowia lypelytica H06	++	
Candida zelanoides H07	+ .	
Candida apis H08	-	
Candida boleticola H09	++	
Candida spandovensis H10	+	
Candida spandovensis H11	+	
Candida tropicalis H12	+	
Candida tropicalis H13	++	
Rhodotorula glutinis H14	++	
Rhodotorula bogoriensis H15	++	
Rhodotorula bogoriensis H16	-	
Candida acuta H17	**	
Candida acuta H18	-	
Candida acuta H19	+	
Candida acuta H20	+	
Control	-	

Key: + = positive only; ++ = positive and collapsed; -: negative. Control: distilled water.

Oil spreading or displacement potential of the yeast isolates

The twenty yeast isolates were also screened for their potential to displace and spread crude oil. Of this number, 11 isolates (55 %) were able to displace the oil. The diameter of oil spread ranged from 1.2 cm to 6.2 cm within 5 to 35 seconds (Table 2). It was observed that Saccharomyces cerevisiae H02, Candida boleticola H09 and Rhodotorula bogoriensis H15 had the highest diameter

of displacement of 6.2, 5.3 and 4.7 respectively within 10 seconds, meaning that they had strong ability to displace crude oil.

Emulsification potential of the yeast isolates

The emulsification capacities of the yeasts were tested and it was observed that the organisms had varying emulsifying capacities which ranged from 6.67 % to 33.33 % (Table 3). Twelve (60 %) of the

^{*}Code assigned to yeasts in our laboratory.

strasburgensis, (Pichia Saccharomyces cerevisiae H03, Candida zelanoides H07, Candida apis H08, Candida spandovensis H10, Candida spandovensis H11, Candida tropicalis Rhodotorula glutinis H14. Rhodotorula bogoriensis H16, Candida acuta H17, Candida acuta H18, and Candida acuta H20) had no emulsifying capacity, 5 (25 %) of the isolates (Yarrowia lipolytica H06, Candida acuta Cryptococcus laurentii H19. Cryptococcus skinneri H05, and Candida

tropicalis H13) had very emulsification indexes of 6.67 %, 8.11 %. % and 16.67 % 16.67 %, 16.67 (15 9/0) respectively, 3 (Saccharomyces cerevisiae H02, Candida and Rhodotorula boleticola H09bogoriensis H15) had emulsification indexes of 33.33 %, 27.33 % and 24.20 % which is considered moderate. However none of the isolates showed percentage emulsification that was significantly high (50 % to 100 %).

Table 2: Diameter and time of displacement caused by yeast isolates

	Table 2: Diameter and time of di	Diameter (cm)	Time (s)	Interpretation
	Coded yeast isolates			
	Laminives cereviside 1102	5.3	7	
53	- Jida holeficola NU9	5.3 4.7	10	
	pladatarula hogoriensis n 13	4.5	12	•
	Pichia strasburgensis H01	4.5		·
	Cryptococcus laurentii H04	3.8	35	
	Saccharomyces cerevisiae H03	3.5	20	Positive
	Candida acuta H19	3.2	34	
	Yarrowia lypolytica H06	2.7	13	
	Cryptococcus skinneri H05	2.3	9	1 Ba
	Candida zelanoides H07	1.2	7	
	Candida tropicalis H13	1.2	16	
	Candida apis HO8		= "	
	Candida spandovensis H10	-	, =	
	Candida spandovensis H11		-	
	Candida tropicalis H12	-	-	
	Rhodotorula glutinis H14			Negative
	Rhodotorula bogoriensis H16	· -	-	- Tregative
	Candida acuta H17	-	-	
	Candida acuta H18		The same of the first	
1	Candida acuta H20	. • · · · · · ·	-	= 1
(Control A		-	
(Control B		5	
		* * *		

Key: Control A=Distilled water; Negative=diameter less than 4.5 and occurred after 30 seconds; Control R=Story B=Sterile broth; Positive=diameter from 4.5 and above and occurred within 30 seconds; cm=centimeter;

Note: The results are arranged in ascending order of displacement with time.

^{*}Code assigned to yeasts in our laboratory

A CONTRACT OF THE PROPERTY OF

Manual Ma		THE STREET, ST
	0.4003	-
List and a rest of the contract of the contrac	38 33 4 3 27	
State and the second of the se	0±00%	
ASS CONTRACTOR SECURITY	166,70	
Contraction of the High	256743935	
5-20 10 12 18 W	のの「土) 240	
Canada do arestas 1907	040CD	
C23222473 373	€00°±0	
स्मि बंगायर वर्ष व्यक्ति	27.33±2.03 ^{bc}	
Caralle sperioterans H10	0±0.00°	
Decid quedivers H11	0 ± 0.00°	
Centile proposeds H12	0 ± 0 CO2	
Certify Terrise is H13	16.67±1.14°	
Sharmagares H14	0+0 002	
Ebaltarale arganiensis H15	24.20±0.875	
E CONTROL OF THE HIG	0+0.00°	
Contracto H17	0+0.004	
Security H18	0+0.002	
Carrier active H19	0±0.00*	
Caradical arcita H20	8.11±0.45°	
Commo	0 + 0.002	
and with a single and a single	2_0.00	

Key: $9.5E_{24}$ =percentage emulsification index; control = distilled water; \pm Se= standard error; the attached letters signifies significant difference (p<0.05). Numbers bearing same letters within rows are not significantly different.

DISCUSSION

Saravana and Vijayakumar (2012) screened 243 and 802 isolates from oil contaminated soil samples using oil drop cellapse for biosurfactant production and only 10 (4.12 %) and 40 (4.99 %) respectively gave positive result. These differences in results may be attributed to the physiological characteristics of the organisms as well as their genetic and composition. molecular researchers (Satpute et al., 2008; Thavasi, et al., 2010; Jain et al., 2011; Tarango et al, 2012; Ibrahim et al., 2013; Pereira et al, 2013; Padmapriya et al., 2013) have proved the oil drop collapse method to be highly sensitive, very effective, and reliable in identification of potent biosurfactant producers

In a study conducted by Chandran and Das (2011), biosurfactants produced by *Rhodotorula muciliginosa* and *Candida rugosa* could effectively emulsify (86 % and 78 %) diesel oil respectively. Other researchers have reported similar results (Chander *et al.*, 2012; Padmapriya *et al.*, 2013).

CONCLUSIONS

Saccharomyces cerevisiae H02, Candida boleticola H09 and Rhodotorula bogoriensis H15 were considered efficient potential candidates for biosurfactants production

^{*}Code assigned to yeasts in our laboratory

REFERENCES

- Chandran, P. and Das, N. (2011).
 Characterization of sophorolipid biosurfactants produced by yeast species grown on diesel oil.

 International Journal of Science and Nature, 2(1), 63-71.
- Chander, C. R. S., Lohitnath, T., Kumar, D. J. M. and Kalaichelvan, P. T. (2012). Production and characterization of biosurfactant from bacillus subtilis MTCC441 and its evaluation to use as bioemulsifier for food biopreservative. Advances in Applied Science Research, 3(3), 1827-1831.
- Hua, Z., Chen, J., Lun, S. and Wang, X. (2003). Influence of biosurfactants produced by
- Candida antarctica on surface properties of microorganism and biodegradation of *n*-alkanes.

 Water Resources, 37, 4143-4150.
- Ibrahim, M. L., Ijah, U. J. J., Manga, S. B., Bilbis, L. S. and Umar, S. (2013). Production and partial characterization of biosurfactant produced by crude oil degrading bacteria.

 Biodeterioration and Biodegradation, 81, 28-34.
- Jain, R. M., Mody, K., Mishra, A. and Jha, B. (2011). Physicochemical characterization of
- biosurfactant and its potential to remove oil from soil and cotton cloth. Carbohydrate Polymers, 10, 10-16.
- Luna, J. M., Rufinho, R. D., Sarubbo, L. A. and Campos-Takaki, G. M. (2013).

- Characterization, surface properties and biological activity of a biosurfactant produced from industrial waste by *Candida Sphearica* ucp0995 for application in the petroleum industry. *Colloids Surface Biointerfaces*, 102, 202-209.
- Padmapriya, B., Suganthi, S. and Anishya, R.S. (2013). Screening, optimization and production of biosurfactants by *Candida* Species isolated from oil polluted soils. *American-Eurasian Journal of Agriculture and Environmental Science*, 13(2), 227-233.
- Pereira, F. B., Gudina, E. J., Costa, R., Vitorino, R., Teixeira, J. A., Coutinho, A. P. and Rodrigues, L. R. (2013). Optimization and characterization of biosurfactant production by Bacillus subtilis isolates. *Fuel*, 111(3), 259-268.
- Rodrigues, L. R., Teixeira, J. A., Van Der Mei, H. C. and Oliveira, R. (2006). Physiochemical and functional characterization of a biosurfactant produced by *Lactococcus lactis*. *Colloid Surface Bioinformatics*, 49, 79-86.
- Saravanan, V. and Vijayakumar, S. (2012).
 Isolation and screening of biosurfactant producing microorganisms from oil contaminated soil. Journal of academic and industrial resources, 1(5), 264-268.
- Satpute, S. K., Bhawsar, B. D., Dhakephalkar, P. K. and Chopade, B. A. (2008). Assessment of

- different screening methods for selecting biosurfactant producing marine bacteria. *Indian Journal of Marine Science*, 37, 243-250.
- Tarango, O. A. L., Moorillon, N. G. V., Casarrubias, B. M. L., Chavira, R. B. E.. and Borunda, O. E. (2012). Isolation and characterization of biosurfactant producing bacteria. *Environmental Engineering and Management Journal*, 11(3), 1-12.
- Thavasi, R., Jayalakshmi, S., Balasubramanian, T. and Ibrahim, M. B. (2009). Production and

- characterization of a glycolopid biosurfactant from Bacillus megaterium using economically cheaper sources. World Journal of Microbiology and Biotechnology, 24(7), 917-925.
- Youssef, H. N., Duncan, K. E., Nagle, D. P.,
 Savage, K. N., Knapp, R. M. and
 McInerney, M. J. (2004)
 Comparison of method to detect
 biosurfactant production by
 diverse microorganisms. Journal of
 Microbiological Methods, 56, 339347.