



## Comparative Larvicidal Efficacy of Leaf and Stem Extract of *Jatropha Curcas* against *Culex Pipiens Pipiens*

Olayemi I K<sup>1</sup>, Busari J<sup>1</sup>, Adeniyi K A<sup>1</sup> and Ukubuiwe A C<sup>1\*</sup>

<sup>1\*</sup>Applied Entomology and Parasitology Research Unit, Department of Biological Sciences, Federal University of Technology, Minna, Nigeria.

\*For correspondence e-mail: azubuike383@gmail.com

Article Info: Received 22 July 2014 Revised: 28 Aug 2014 Accepted 09 Sep 2014

### ABSTRACT

The present study aimed to investigate the larviciding efficacy of stems and leaves extract of *Jatropha curcas*, against larval stage of the *Culex pipiens pipiens* mosquito. Methanolic extracts of the plant material were prepared and bio-assayed against the larvae as per standard World Health Organization's (WHO) procedures. The larvae were exposed to a series of concentrations of the extracts ranging from 0.25 - 3.75 mg/l; and mortality recorded post-24 hours exposure. The results showed that larval mortality increased significantly ( $P < 0.05$ ) with increasing in extracts concentration. Mortality ranged from  $2.00 \pm 1.40\%$  in the lowest leaf-extract concentration (i.e., 0.25 mg/l) to 96% in the highest (3.75 mg/l). Generally, mortalities were relatively higher among larvae exposed to stem extract than that of leaf. Probit regression analysis of the mortality data gave  $LC_{50}$  and  $LC_{90}$  of the leaf extract as 2.65 and 3.45 mg/l respectively; while those of the stem were 2.33 and 3.25 mg/l respectively. These results indicate that the stem extract of *J. curcas*, in particular, is a potential source of a mosquito larviciding lead-agent, and its further exploitation should fast-track the development of a cost-effective larvicide for integrated mosquito vector control.

**Keywords:** Bio-assay, Extracts, Larva, Lethal Concentration, Mortality and larviciding efficacy.

### 1. INTRODUCTION

Mosquitoes are insect vectors of human diseases including, Malaria, Filariasis, Dengue fever, Japanese encephalitis, etc. [1]. These diseases cause several million cases of morbidity and mortality per annum, in their various regions of distribution [2]. For example, Filariasis transmitted by the mosquito genus *Culex*, threatens the health of over a billion people world-wide with about 2.5 million deaths reported yearly [3]. In addition, the *Culex* mosquitoes constitute the most abundant Culicids in urban areas [4]. This development has made the control of *Culex*

mosquitoes imperative. Yet, the conventional mosquito control strategy in sub-Saharan Africa (i.e., the use of insecticide-treated bed nets), is targeted at taking advantage of the behaviour of anopheline mosquitoes that bite mostly indoors and late at night [5]. This vector control strategy is not effective against *Culex* mosquitoes that bite principally outdoors before people go to sleep under the protective bed-nets. Therefore, larval control measures would be more effective against *Culex* mosquitoes, taking its adult behavior into consideration.

However, the effective mosquito larviciding interventions often involve the use of synthetic chemicals that are expensive, scarce and harmful to the ecosystem [6, 7]. These challenges have called for the search for mosquito larvicides from cheap and readily available sources, especially, natural products of plants origin [8]. To this end, various plant taxonomic families have been screened for insecticidal properties, with encouraging results obtained in many cases [9-11]. Studies have revealed that *Jatropha curcas* (Physic nut) is rich in bio-active phytochemicals that have found applications in medicine [12-14].

Insecticidal efficacies of extracts of *Jatropha curcas* have been demonstrated against a wide range of insect species including, the Tobacco hornworm (*Manduca sexta*), cotton bollworm (*Helicoverpa armigera*), melon Aphid (*Aphis gossypii*), leafhopper (*Enaposcabi guttula*), maize weevil (*Sitophilus zeamays*), etc. [15-17]. However, information on larviciding activities of *J. curcas* against mosquito species in general, *Culex pipiens pipiens* in particular is very scanty. Therefore, this study was carried out to determine the larvicidal efficacies of the extracts of *J. curcas* against the *Culex pipiens pipiens* mosquito. This is against the back drop that *J. curcas* is widely distributed in the Tropics, where its cultivation is highly encouraged for medicinal uses and production of bio-diesel.

## 2. MATERIALS AND METHODS

### 2.1. Source of Plant Materials and Preparation of Extracts

Fresh leaves and stems of *Jatropha curcas* were obtained from plants growing in the field, around the Temporary Campus of Federal University of Technology, Minna, Nigeria. The taxonomic identity of the plant materials was authenticated by a Botanist in the Department of Biological Sciences, Federal University of Technology, Minna, Nigeria; after which voucher specimens of the plant materials were deposited in the Herbarium of the Department.

For preparation of extracts, the plant materials were air-dried at ambient Laboratory conditions. Then, the dried plant materials were pulverized using an electric grinder (Model: WPB80BC), and the powder stored at 28°C until used. Thereafter, 25g of the individual plant material (i.e., leaves or stems) was extracted in 300ml methanol, using Soxhlet apparatus, at 80°C for a period of 16hours [18]. The crude extracts were purified using a rotary vacuum

evaporator, and the stock solutions, subsequently prepared, were stored in a refrigerator at 4°C.

The *Culex pipiens pipiens* mosquitoes used for this study were originally obtained from the wild in Minna, Nigeria, and maintained in the Laboratory of the Department of Biological Sciences, Federal University of Technology, Minna, Nigeria. Laboratory handling and maintenance of the mosquitoes, followed standard mosquito rearing protocols [19].

### 2.2. Preparation of Working Solution of Extracts

The stock solution was serially diluted in ten-fold with distilled water (i.e., 2 ml stock solution to 18 ml solvent), following recommended guidelines [20]. Test concentrations were then obtained by adding a range of 0.25 - 3.75 mg/l (in multiples of 0.50 mg/l), to 100ml of distilled water.

### 2.3. Bioassay of Plant Extracts against *Cx. p. pipiens* Larvae

Larvicidal bioassays were conducted for 24 hours in plastic containers of 250ml capacity. Batches of 25 healthy 4<sup>th</sup> instar larvae of the mosquito were transferred, by means of a strainer, to plastic containers holding 100ml distilled water. The appropriate volume of extract was added to 100ml of water in the test containers, to obtain desired extract concentration.

Four replicates were set up for each test concentration and an equal number of Controls were also set up simultaneously; with distilled water to which only 1ml of the methanol solvent was added. The experiment was carried out at ambient Laboratory conditions of 28.00±1°C, 82.00±2% relative humidity and 12 hr.: 12hr light: darkness photoperiod. Larval mortality was recorded post-24 hours exposure to the extracts. The whole experiment was repeated within 24 hours of termination of the first exercise.

### 2.4. Statistical Analysis

Larval mortality data obtained were subjected to Analysis of Variance (ANOVA) using software ware package SPSS 16 version. Statistical differences between the treatments was determined by Duncan multiple range test (P<0.05). Probit regression analysis [21], was done using Biostat 2012 software for the determination of LC<sub>50</sub> and LC<sub>90</sub> of the different extracts.

### 3. RESULTS

Table 1 shows the concentration-dependent effects of leaf and stem extracts of *J. curcas*, against 4th instar larval stage of *Culex pipiens pipiens* mosquitoes, under Laboratory conditions. For both extracts, mortality rates of the larvae increased significantly ( $P < 0.05$ ) with rising extract concentration. Mortality ranged from  $2.00 \pm 1.40\%$  in the lowest leaf-extract concentration tested (i.e.,  $0.25 \text{ mg/l}$ ) to  $96.00 \pm 0.00\%$  in the highest concentration ( $3.75 \text{ mg/l}$ ). However, the respective percentage ranges were  $0.00 \pm 0.00\%$  and  $100.00 \pm 0.00\%$  for the stem extract. Except for the first three extract concentrations tested (i.e., range =  $0.25 - 1.25 \text{ mg/l}$ ), larval mortalities recorded among mosquitoes exposed to the stem extract were significantly higher than those of leaf-extract.

The lethal concentrations (LC) of the extracts required to kill 50 and 90% (i.e.,  $LC_{50}$  and  $LC_{90}$ , respectively) of the larvae are highlighted in Table 2. The  $LC_{50}$  of the leaf and stem extracts were, respectively,  $2.65$  and  $2.33 \text{ mg/l}$  while, the  $LC_{90}$  values were  $3.45$  and  $3.25 \text{ mg/l}$ , respectively.

### 4. DISCUSSION

The extracts of *J. curcas* bio-assayed in this study, demonstrated significant larvicidal activities against *Cx. p. pipiens*, and such activities were dose-dependent. The concentration-dependent effects of the extracts indicate the presence of mosquito larvicidal bio-active phytochemicals in the leaves and stems of *J. curcas*. This observation supports the results of Rahuman et al. [22], who reported significant larvicidal activities of *J. curcas* against different mosquito species. The larvicidal activities of the extracts may be due to the presence of Flavonoids, Saponins, Glycosides, etc [23], which, according to literature, are highly toxic to mosquito larvae [11, 24, 25]. On the whole, the stem extract elicited much higher mortalities in the larvae, than the leaf-extract. These differential larvicidal activities of the leaf and stem extracts may be due to variations in phytochemical composition and/or concentrations of such phytochemicals in the two organs of *J. curcas*. According to Oseni and Alphonse [26], while Flavonoid and Phenolic phytochemicals were present in the stem, they were absent from the leaf; also, alkaloids occurred in a higher concentration in the stem than leaf of *J. curcas*. These phytochemicals, that are either present only in the stem or occurred there in higher concentrations, have been reported as

responsible for the insecticidal of many plant extracts [11, 27].

**Table 1.** Mean mortality of *Culex pipiens pipiens* mosquito larvae, exposed to varying concentrations of leaf and stem extracts of *J. curcas*

Extract concentration (mg/ml)	Plant material extracted	
	Leaf	Stem
0.00 (Control)	$0.00 \pm 0.00^a$	$0.00 \pm 0.00^a$
0.25	$2.00 \pm 1.40^b$	$0.00 \pm 0.00^a$
0.75	$10.00 \pm 3.46^c$	$6.00 \pm 1.64^b$
1.25	$18.00 \pm 4.68^d$	$18.00 \pm 2.60^c$
1.75	$38.00 \pm 4.88^f$	$47.00 \pm 3.28^d$
2.25	$28.00 \pm 4.52^e$	$55.00 \pm 4.32^d$
2.75	$56.00 \pm 2.24^e$	$75.00 \pm 5.88^e$
3.25	$80.00 \pm 2.70^h$	$85.00 \pm 5.88^f$
3.75	$96.00 \pm 0.00^i$	$100.00 \pm 0.00^e$

<sup>a</sup>Values followed by different superscript alphabets in a column are significantly different at  $P = 0.05$ .

**Table 2.**  $LC_{50}$  and  $LC_{90}$  (mg/ml) of methanolic extract of *Jatropha curcas* against 4<sup>th</sup> instar larval stage of *Culex pipiens pipiens* mosquitoes

Extract	$LC_{50}$ (Confidence Limits)	$LC_{90}$ (Confidence Limits)
<b>Leaf</b>	$2.65$ (2.19 – 2.79)	$3.45$ (3.22 – 3.60)
<b>Stem</b>	$2.33$ (2.14 – 2.51)	$3.25$ (3.08 – 3.37)

### 5. CONCLUSION

The findings of this study have shown that extracts of the stems and leaves of *J. curcas* possess significant larvicidal potentials against the *Cx. p. pipiens* mosquito and, therefore, may be a veritable source of insecticidal lead-agents for mosquito vector control. However, the stem extract appears to be more

promising as a larvicide, due to its richer qualitative and/or quantitative phytochemical diversity. This finding, therefore, suggest that more attention should be focused on the stem extract of *J. curcas*, through fractional bio-assay, for optimum larvicidal efficacy. The results of this study have provided base-line information that should fast-track and guide the development of cost-effective eco-friendly larviciding agents for integrated mosquito vector control.

### Acknowledgement

The authors wish to thank Dr. I. K. Olayemi, Head of Department, Biological Sciences for providing laboratory facilities. The authors wish to thank the Botanist, Mr. Daud Yusuf O for the identification of the plant species. The valuable support from Laboratory technicians and Assistants are gratefully acknowledged.

### Conflict of Interest

The authors declare that they have no conflicts of interest.

### References

1. Gubler DJ (1998). Resurgent vector borne diseases as a global health problem. *Emerg. Infect. Dis*; 4: 442-50.
2. World Health Organisation (2004). *Community participation and tropical disease control in resource poor settings*. TDR/STR/SEB/ST/04.1
3. World Health Organization (2011). Lymphatic filariasis". <http://www.who.int/m-ediacentre/factsheets/fs102/en/> retrieved on 3/11/2011.
4. Samuel T, Arivoli S and Jesudoss M (2011). Larvicidal Efficacy of Vernonia Cinerea (L.) (Asteraceae) Leaf Extracts against the Filarial Vector *Culex quinquefasciatus* say (Diptera: Culicidae), *Journal of Biopesticides*; 4(1): 37 – 42.
5. Olayemi IK and Ande AT (2008). Nocturnal biting behaviour of anopheline mosquitoes (Diptera: Culicidae) in Ilorin, Nigeria. *Nigerian Journal of Applied and Experimental Biology*; 9(2): 83-89.
6. Casida JE and Quistad GB (2000). Insecticide targets: Learning to keep up with resistance and changing concepts of safety. *Agric. Chem Biotechnol*; 43: 185-191.
7. Devine GJ and Furlong MJ (2007). Insecticide use: Contexts and ecological

- consequences. *Agr. Human Values*; 24: 281-305.
8. Pitasawat B, Choochote W, Kanjanapothi D, Panthong A, Jitpakdi A and Chaithong U (1998). Screening for larvicidal activity of ten carminative plants. *Southeast Asian Journal of Tropical Medicine*; 29:660-672.
9. Ciccia G, Cussio J and Mongelli EJ (2000). Insecticidal Activity of some Medicinal Plants against *Aedes aegypti* Mosquito Larvae of South American. *Journal of Ethnopharmacology*; 72: 185-189.
10. Narumon K, Yuwadee T, Yupha R and Chamnarn A (2005). Screening for Larvicidal Activity in some Thai Plants against four Mosquito Vector Species. *Journal of Medical Entomology*; 36(6): 14-16.
11. Shallan EA, Canyon D, Younes MWF, Abdel-Wahab H and Mansour A (2005). A review of botanical phytochemicals with mosquitocidal potential. <http://dx.doi.org/10.1016/j.envint.2005.03.003>
12. Igbinosa OO, Igbinosa EO and Aiyegoro OA (2009). Antimicrobial Activity and phytochemical Screening of Stem Bark Extracts from *Jatropha curcas* (Linn). *African Journal of Pharmacy and Pharmacology*; 3(2): 058–062
13. Rug M and Ruppel A (2000). Toxic Activities of the Plant *Jatropha curcas* against Intermediate Snail Hosts and Larvae of Schistosomes. *Journal of Tropical Medicine and International Health*; 5: 423–430.
14. Thomas R, Sah NK and Sharma PB (2008). Therapeutic Biology of *Jatropha curcas*: a Mini Review. *Current Pharmaceutical Biotechnology*; 9: 315-324.
15. Selke SS, Jadhav LD and Salunkhe GN (1985). Oviposition and Adult Repellent Action of some Vegetables Oils/Extracts against Potato Tuber Moth. *Journal of Maharashtra Agricultural*; 32(7): 67–72.
16. Sauerwein M, Sporer F and Wink M (1993). Insect-Toxicity of Phorbol Esters from *Jatropha curcas* Seed Oil. *Planta Medica*; 59: 6867–7233.
17. Wink M, Koschmieder C, Sauerwein M and Sporer F (1997). Phorbol Esters of *Jatropha curcas* – Biological Activities and Potential Applications. In: Gubitzi GM, Mittelbach M Trabi M (eds) *Biofuel and Industrial Products from Jatropha curcas Indian Journal of Science*; 20: 12–16.

18. Koyel MH, Papiya G and Goutam C (2011). Evaluation of Target Specific Larvicidal Activity of *Triphonium trilobatum* against *Culex quinquefasciatus*, *Asian Pacific Journal of Tropical Biomedicine*; 3: 199–203.
19. Olayemi IK and Ande AT (2009). Life Table Analysis of *Anopheles gambiae* (Diptera: Culicidae) in Relation to Malaria Transmission. *Journal of Vector Borne Diseases*; 46: 295-298.
20. World Health Organization (2005). Malaria vector control and personal protection. 936<sup>th</sup> technical report series, 2005, World Health Organization.
21. Finney DJ (1971). *Probit analysis*, Cambridge University Press, London.
22. Rahuman AA, Gopalakrishnan G, Venkatesan P and Geetha K (2007). Larvicidal Activity of some Euphorbiaceae Plant Extracts against *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitology Research*; 102: 867-873.
24. Chapagain BP, Saharan V and Wiesman Z (2008). Larvicidal Activity of Saponins from *Balanites aegyptiaca* Callus against *Aedes aegypti* Mosquito. *Bioresource Technology*; 99(5):1165-1168.
25. Quevedo R, Baquero E and Quinones ML (2011). 1-Phenylisoquinoline larvicidal activity against *Culex quinquefasciatus*. *Natural Product Research*; DOI:10.1080/14786419.2011.560846
26. Oseni LA and Alphonse PK (2011). Comparison of Antimicrobial Properties of Solvent Extracts of Different Parts of *Jatropha curcas*, *International Journal of Pharmacy and Pharmacology*; 1(3): 117–123.
27. Sukumar K, Perich MJ and Boobar IR (1991). Botanical derivatives in mosquito control: a review. *Journal of American Mosquito Control Association*; 7(2): 10-37.