

Antimicrobial and Free Radical Scavenging Potentials of *N*-Hexane and Ethyl Acetate Fractions of *Phyllanthus Fraternus*

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1 ABSTRACT

2 The genus *Phyllanthus* (Phyllantaceae) is widely used in the african system of traditional medicine and
 3 is reported to have various biological activities. In this study, antimicrobial and antioxidant activities of *n*-
 4 hexane and ethyl acetate fractions of *Phyllanthus fraternus* leaves were investigated. The antimicrobial
 5 screening was carried out against *Staphylococcus aureus*, *Escherichia coli*, *Bacillus subtilis*,
 6 *Pseudomonas aeruginosa*, *Salmonella typhi* and *Klebsiella pneumoniae*, using Agar-well diffusion
 7 method. The antioxidant activity was carried out using DPPH free radical scavenging capacity. The
 8 results show that fractions of *Phyllanthus fraternus* leaves have DPPH radical scavenging activities with
 9 IC₅₀ value of 263.53 mg/mL and 143.56 mg/mL for *n*-hexane and ethyl acetate fractions respectively.
 10 For *n*-hexane fraction, the MICs of the extract were; 80 mg/mL against *K. pneumoniae* and *S.*
 11 *aureus*, 120 mg/mL against *P. aeruginosa* and *S. typhi*, and 160 mg/mL against *E. Coli*. However, ethyl
 12 acetate fraction had MICs of 80 mg/mL against all test organisms except *S. aureus* (40 mg/mL). The *n*-
 13 hexane and ethyl acetate fractions of *Phyllanthus fraternus* leaves exhibited considerable antioxidant
 14 and antimicrobial properties, with ethyl acetate fraction been the most potent. This plant extract can be
 15 regarded as promising resource for antimicrobial and antioxidant drugs.

16 **Keywords:** Antioxidant; Antimicrobial; *Phyllanthus fraternus*; *n*-hexane, ethyl acetate fractions.

17 INTRODUCTION

18 Africa is endowed with large amounts of
 19 medicinal plants used for therapeutic
 20 intervention (Bashir *et al.*, 2015; Lawal *et al.*,
 21 2015; Lawal *et al.*, 2016a). The importance of
 22 plants in medicine remains of greater relevance
 23 with the current global shift to obtain drugs from
 24 plants sources, as a result of which attention
 25 has been given to the medicinal value of herbal
 26 remedies for safety, efficacy and economy
 27 (Adebayo *et al.*, 2009). Plants constitute an
 28 important source of active ingredients which
 29 differ widely in terms of structure and
 30 therapeutic properties (Lawal *et al.*, 2016b).
 31 The continued investigation into the secondary
 32 plant metabolites for anti-infective properties
 33 has gained importance in recent years because
 34 of the alarming increase in resistance of
 35 pathogenic microorganisms to existing
 36 antibiotics. For instance, the emergence and
 37 spread of *Salmonella* resistance to many
 38 commonly used antibiotics (Ciprofloxacin,

40 Ampicillin, Chloromphenicol, Amoxicillin) has
 41 been a subject of international concern (Tsobou
 42 *et al.*, 2015).

43
 44 The recent growth in knowledge of free radicals
 45 and Reactive Oxygen Species (ROS) in
 46 biological systems is causing a medical
 47 revolution that promises a new age of health
 48 (Tsado *et al.*, 2016). Free radicals are highly
 49 reactive molecules generated during oxidation
 50 reactions which in turn initiate chain reactions
 51 resulting in to cellular damage (Lawal *et al.*,
 52 2015b). There is substantial evidence
 53 implicating free radicals especially reactive
 54 oxygen species (ROS) in the etiology of more
 55 than one hundred degenerative disorders in
 56 humans including, arthritis, atherosclerosis,
 57 ischemia and reperfusion injury of many
 58 tissues, gastritis, diabetics, central nervous
 59 system injury, acquired immunodeficiency
 60 syndrome (AIDS) and cancer (Lawal *et al.*,
 61 2016a)

62
63 Reports abound on the antioxidant activities of
64 phytochemical constituents of medicinal plants
65 (e.g. polyphenols, carotenoids, flavonoids,
66 phenolics, vitamins C and E). These
67 phytochemicals act as antioxidants by
68 preventing damages to cell membrane due to
69 cellular oxidative processes that may result in
70 diseases (Soni *et al.*, 2015). For instance,
71 natural polyphenols from plants have been
72 found to exert their beneficial effect by
73 removing free radicals, chelating metal catalyst,
74 activating antioxidant enzymes, etc (Lawal *et al.*, 2016a).
75 *Phyllanthus fraternus* G.L.Webster
76 (*Phyllanthaceae*) is widely distributed in most
77 tropical and subtropical countries, and have
78 long been extensively used in folk medicine in
79 Africa and most other countries for thousands
80 of years in the treatment of a broad spectrum of
81 diseases, such as disturbances of the kidney
82 and urinary bladder, intestinal infections,
83 diabetes, and the hepatitis B virus (Manjulatha
84 *et al.*, 2008). The present study sought to
85 evaluate antimicrobial and antioxidant activities
86 of *n*-hexane and ethyl acetate fractions of
87 *phyllanthus fraternus*.

89

90 MATERIALS AND METHODS

91 Chemicals

92 DPPH (2,2-diphenyl-1-picrylhydrazyl) and
93 solvents use were obtained from Sigma-Aldrich
94 (Steinhein-Germany), All solvents used for
95 extraction were of analytical grade.

96

97 Plant Collection

98 Freshly harvested *Phyllanthus fraternus* leaves
99 were procured from Bosso, area of Minna,
100 Niger State, Nigeria. The plant was
101 authenticated by a botanist at National Institute
102 of Pharmaceutical Research and Development,
103 Abuja, Nigeria.

104

105 Sources of Microorganisms

106 Pure isolates of *K. pneumoniae*, *S. aureus*, *P.*
107 *aeruginosa*, *E.coli* and *S. typhi* were procured
108 from Microbiology Unit, Faculty of Life Sciences
109 Federal University of Technology, Minna,
110 Nigeria. Biochemical test and Gram staining

111 test were used to confirm the identity of the
112 organism.

113

114 Extraction of plant Materials

115 Fresh leaves of *Phyllanthus fraternus* were
116 grounded using a grinder mill. Exactly 200 g of
117 the powdered plant was extracted with 600ml of
118 methanol. The resulting extract was
119 concentrated using rotary evaporator. The
120 methanol extract was partitioned between *n*-
121 hexane and water. The aqueous layer was
122 further fractionated using different solvents in
123 increasing order of polarity: *n*-hexane,
124 chloroform and ethyl acetate. The fractions
125 were collected and concentrated using rotary
126 evaporator (Resona, Germany). The
127 concentrated fractions were investigated for
128 antimicrobial and antioxidant activities

129

130 Assay for antibacterial activity

131 Stock cultures were maintained at 4°C on
132 nutrient agar (HiMedia) slants. Active cultures
133 for experiments were prepared by transferring a
134 loopful of culture to 10 mL of nutrient broth
135 (HiMedia) and incubated at 37 °C for 24 hours
136 for bacterial proliferation (Jayaraman *et al.*,
137 2008). Antibacterial activity of *n*-hexane fraction
138 of *Phyllanthus fraternus* leaves was carried out
139 using agar-well diffusion method as described
140 by Jayaraman *et al.*, (2008), using Ciprofloxacin
141 (40 µg/mL) as standard drug. Minimum
142 inhibitory concentration (MIC) and minimum
143 bactericidal concentration (MBC) were
144 determined by tube dilution method for each of
145 the test organism in triplicates.

146

147 Estimation of 2,2-diphenyl-1-picrylhydrazyl (DPPH) Radical Scavenging Activity

148 The free radical scavenging activity of the *n*-
149 hexane fraction was assayed using 2,2-
150 diphenyl-1-picrylhydrazyl (DPPH) free radical
151 was determined (Szabo *et al.*, 2007).

152

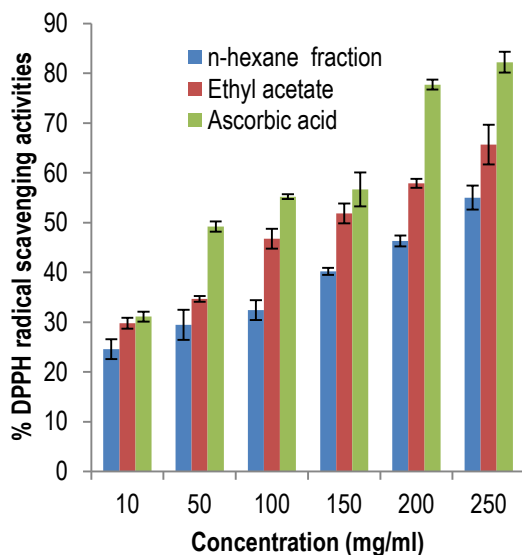
153 Statistical Analysis

154 All the experiments were carried out in triplicate
155 and data obtained from the study were
156 subjected to analysis of variance using
157 statistical package for Social Science (SPSS)
158 version 21 and presented as means ± SE of the
159 mean.

160

161 **RESULTS AND DISCUSSION**

162 Figure 1 shows the results of scavenging
163 radical ability of *n*-hexane and ethyl acetate
164 fractions of *Phyllanthus fraternus* at various
165 concentrations in comparison with same doses
166 of ascorbic acid. The extract was found to exert
167 antioxidants effect in DPPH radical scavenging
168 assay with IC₅₀ value of 263.53 mg/ml and
169 143.56mg/ml for *n*-hexane and ethyl acetate
170 fractions respectively (Figure 2). The decrease
171 in absorbance of DPPH caused by *n*-hexane
172 fraction of *Phyllanthus fraternus* was due to the
173 reaction between antioxidant molecules and
174 radicals, which results in the scavenging of the
175 radical by hydrogen donation.
176



177
178 **Figure 1:** DPPH radical scavenging activities of
179 *n*-hexane and ethyl acetate fractions of
180 *Phyllanthus fraternus* leaves.
181

182 Many antioxidants compounds are present in
183 natural products. Flavonoids are phenolic
184 compounds with important functions in
185 scavenging free radicals and thus play vital
186 roles in preventing oxidative stress associated
187 disorder (Nahak and Sahu, 2010). However, the
188 IC₅₀ value recorded in this study were higher
189 than IC₅₀ values of 41.05, 17.52 and 32.66
190 µg/mL reported for crude methanol fruit extracts
191 of *Phyllanthus acidus*, *Phyllanthus emblica* and
192 *Phyllanthus fraternus*, respectively (Manjulatha
193 *et al.*, 2014). The quality and quantity of
194 bioactive antioxidative agents in plants vary
195 with the plant species, part of the plant used as

196 well as the solvents used in the extraction
197 process (Lawal *et al.*, 2014). Thus the higher
198 IC₅₀ value observed for fractions of *Phyllanthus*
199 *fraternus* leaves could be attributed to species
200 differences and part of the plant used.

201 The antimicrobial effects of plant extracts have
202 been the subject of many studies during the last
203 three decades (Tsobou *et al.*, 2015). Recently,
204 many antimicrobial screening evaluation studies
205 have been published based on traditional
206 Chinese, African and Asian use of extractives
207 that are plant-based (Suffredim *et al.*, 2004). In
208 the present study, the results of antibacterial
209 property of *n*-hexane and ethyl acetate fractions
210 of *Phyllanthus fraternus* leaves (Tables 2 and 3
211 respectively) against tested organisms varied
212 depending on bacteria tested and concentration
213 (Ravikumar *et al.*, 2007; Rajasekharan and
214 George, 2010).
215

216 Increase in the concentration of *n*-hexane and
217 ethyl acetate fractions of *Phyllanthus fraternus*
218 resulted in corresponding increase in the zones
219 of inhibition. This linear relationship between
220 the concentrations of extracts and zones of
221 inhibition could be that the higher concentration
222 of extracts causes a higher diffusion of the
223 substances in the nutrient agar (Tsado *et al.*,
224 2016) The extracts were more active with
225 greater zone of inhibition observed at
226 concentrations of 120 and 160 mg/mL
227 suggesting a dose dependent growth
228 inhibition (Tsado *et al.*, 2016). Antimicrobial
229 activities of most medicinal plants are
230 attributed to the presence of bioactive
231 phytochemicals (Rice-Evans *et al.*, 1995). The
232 methanol extract of *Phyllanthus fraternus*
233 leaves have been reported to contain tannins,
234 saponins, alkaloids, anthraquinones and resins.
235 These phytochemicals reported to offer great
236 pharmacological activities both in traditional and
237 orthodox medicine could be responsible for the
238 enhanced activity of the fractions of *Phyllanthus*
239 *fraternus* leaves as shown in Tables 1 and 2.
240 For *n*-hexane fraction, the MICs of the extract
241 were 80 mg/mL against *K. pneumoniae* and *S.*
242 *aureus*, 120 mg/mL against *P. aeruginosa* and
243 *S. typhi*, and 160 mg/mL against *E. Coli*. The
244 ethyl acetate fraction had MIC of 80 mg/mL
245

246 against all test organisms except for *S. aureus* 251 lower on all test organism compare to zone of
 247 where the MIC was 40 mg/mL (see Table 3). 252 inhibitions demonstrated by standard antibiotics
 248 However, despite the higher zones of inhibition 253 drugs (ciprofloxacin).
 249 demonstrated by fractions of *Phyllanthus* 254
 250 *fraternus* leaves, the zones of inhibitions were

255

256 **Table 1:** Zones of inhibition of *n*-hexane fraction of *Phyllanthus fraternus* leaves against some
 257 pathogenic organism

Concen. (mg/mL)	<i>E. coli</i>	<i>K. pneumoniae</i>	<i>S. aureus</i>	<i>P. aeruginosa</i>	<i>S. typhi</i>
Zone of inhibition (mm)					
40	-	-	-	-	-
80	-	12.00±0.50	14.00±0.10	-	-
120	-	16.00±0.50	21.00±0.10	12.00±1.00	16.00±0.50
160	18.00±0.00	-	11.00±0.50	15.00±0.50	16.00±0.10
180	20.00±0.50	-	11.00±0.05	23.00±0.60	20.00±0.05
Control (40 µg/mL)	18.00±0.00	28.00±0.00	32.00±0.56	28.50±0.40	19.00±0.55

258 Data represent means ± SEM of triplicate determination.

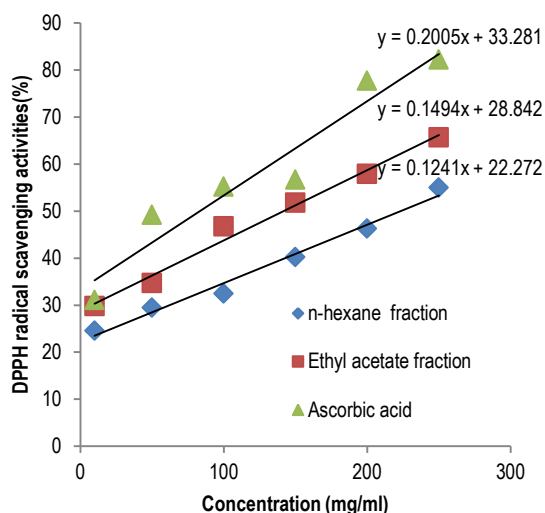
259

260 **Table 2:** Zones of inhibition of ethyl acetate fraction of *Phyllanthus fraternus* leaves against some
 261 pathogenic organism

Concen. (mg/mL)	<i>E. coli</i>	<i>K. pneumoniae</i>	<i>S. aureus</i>	<i>P. aeruginosa</i>	<i>S. typhi</i>
Zone of inhibition (mm)					
40	-	-	5.03±0.70	-	-
80	5.89±0.59	12.45±0.46	9.47±0.38	8.90±0.89	9.08±0.90
120	7.45±0.90	11.45±0.21	9.92±0.36	12.35±0.79	12.30±0.52
160	11.80±0.46	19.90±0.05	13.90±0.55	13.79±0.29	17.08±0.79
180	13.89±0.97	24.79±0.55	16.05±0.50	19.56±0.89	22.47±0.92
Control (40 mg/mL)	18.00±0.00	28.00±0.00	32.00±0.56	28.50±0.40	19.00±0.55

262 Data represent means ± SEM of triplicate determination.

263



264
265 **Figure 2:** DPPH radical scavenging assay for
266 determination of IC₅₀ of *n*-hexane and ethyl
267 acetate fractions of *Phyllanthus fraternus*
268 leaves.

269
270 **Table 3:** Minimal inhibitory concentrations
271 (MIC) of *n*-hexane and ethyl acetate fractions of
272 *Phyllanthus fraternus* leaves against some
273 pathogenic organisms

Test organisms	MIC (mg/mL)	
	N hexane	Ethyl acetate
<i>E. coli</i>	160	80
<i>K. pneumoniae</i>	80	80
<i>S. aureus</i>	80	40
<i>P. aeruginosa</i>	120	80
<i>S. typhi</i>	120	80

274
275 **CONCLUSION**

276 The *n*-hexane and ethyl acetate fractions of
277 *Phyllanthus fraternus* leaves exhibited
278 antioxidant and antimicrobial properties with
279 ethyl acetate fraction been the most potent. The
280 observed activities sports the ethno medicinal
281 use of this plant. The plant extracts could be
282 regarded as a promising source for
283 antimicrobial and antioxidant agents.

284
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